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TwistDx™

Unwind DNA's possibilities

TwistAmp™ Liquid Basic

Quick Guide

Part number: INLQBAS

Revision 4

TwistDx™

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TwistAmp™ Liquid Basic Quick Guide

Please see **instruction and assay design manuals** at twistdx.co.uk for information regarding components and storage, assay design, and detailed use.

Instructions are based on 50 µl reaction volumes; if using a different volume, quantities should be adjusted appropriately.

Primer screen set-up (single-plex)¹

1. Add 2.4 µl of each primer at 10µM concentration to 0.2 ml PCR tubes.
2. Prepare a pre-master mix (per reaction) in the order below:

2x Reaction Buffer	25 µl
dNTPs ² + water ³ to	9.2 µl
10x Basic E-mix	5 µl

Vortex and spin briefly.
3. To the pre-master mix, add 2.5 µl 20x Core Reaction Mix⁴ (per reaction) to tube lid. Mix by 10x full inversions and spin briefly. Master mix is now complete⁵. Pipette mix before use.

4. Add 41.7 µl³ of master mix to primers prepared in tubes (step 1) and pipette mix.
5. Add 2.5 µl of 280mM MgOAc (supplied) and 1 µl template to tube lids³. DNA and MgOAc should be kept separate in the tube lid prior to spindown. Spin in MgOAc/template and mix well (6x inversions) to start reaction. Spin briefly.

Warning: RPA reactions start as soon as MgOAc is added.

6. Incubate at 37-42°C for 20-40 minutes. For low template copies, remove strip after 4 mins, mix by 6x full inversions and spin briefly, replace in heating device.
7. After step 6, clean amplicons before running on an agarose gel.

Warning: Opening tubes post amplification will risk contamination of work surfaces with amplicon. Ensure appropriate control measures are taken.

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Template screen set-up (single-plex)¹

1. Prepare a primer pre-master mix (per reaction) in the following order:

2x Reaction Buffer	25 µl
dNTPs ² + water ³ to	9.2 µl
10x Basic E-mix	5 µl
Primer A (10µM)	2.4 µl
Primer B (10µM)	2.4 µl

Vortex and spin briefly.
2. Add 2.5 µl 20x Core Reaction Mix⁴ (per reaction) to tube lid. Mix by 10x full inversions and spin briefly. Master mix is now complete⁵. Pipette mix before use.
3. Add 46.5 µl³ master mix to 0.2 ml PCR tubes.
4. Add 2.5 µl of 280mM MgOAc and 1 µl template to tube lid³. DNA and MgOAc should be kept separate in the tube lid prior to spin-down. Spin in MgOAc/template, mix well (6x inversions) to start reaction. Spin briefly.

Warning: RPA reactions start as soon as MgOAc is added.

5. Incubate at 37-42°C for 20-40 minutes. For low template copies, remove strip after 4 mins, mix by 6x full inversions and spin briefly, replace in heating device.
6. After step 5, clean amplicons before running on an agarose gel.

Warning: Opening tubes post amplification will risk contamination of work surfaces with amplicon. Ensure appropriate control measures are taken.

- ¹ See manual for multiplexing.
- ² Suggested final concentration of 1.8mM (total) dNTPs. Optimisation is recommended.
- ³ Volumes should be adjusted if adding more/less template and/or MgOAc.
- ⁴ Warm to room temperature and pipette mix slowly to ensure homogeneity.
- ⁵ Master mix may appear cloudy, this is normal.

TwistDX
Liquid Basic QG

Size:
8.268 in x 5.827 in



PMS 185 C
Red



Black



PMS 7541 C
Gray 1 - 10%

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