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TwistAmp™ Basic Kit

Quick Guide

Part number: INTABAS

Revision 3



TwistAmp™ Basic Kit Quick Guide

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Please see instruction and assay design manuals at twistdx.co.uk for information regarding kit components and storage, assay design, detailed use and multiplexing.

Set-up (single-plex)

1. Prepare reaction mix in 1.5ml tube:

Primer A (10μM) $2.4 \mu l$ Primer B (10μM) $2.4 \mu l$ Primer Free Rehydration buffer $29.5 \mu l$ Template and water to $13.2 \mu l$ (Total volume $47.5 \mu l$)Vortex and spin briefly.

- 2. Add reaction mix to a TwistAmp® Basic reaction. Pipette to mix.
- 3. Add 2.5 µl of 280mM Magnesium Acetate (MgOAc) (supplied) and mix well to start reaction.

Note: RPA reactions start as soon as MgOAc is added.

- 4. Incubate at 39°C for 20 minutes. For low template copy number, remove strip after 4 minutes, vortex and spin briefly, replace in heating device.
- 5. After 20 minutes, clean amplicons before running on agarose gels.

Note: If tubes are opened after amplification there is a great risk of contamination of work surfaces with amplicon. Ensure that appropriate avoidance measures are taken.

Kit Positive Control Set-up (single-plex)

1. Prepare reaction mix in 1.5ml tube:

Positive control primer mix $8 \mu l$ Primer Free Rehydration buffer $29.5 \mu l$ Positive control DNA template $1 \mu l$ Water $9 \mu l$ (Total volume $47.5 \mu l$)

- Vortex and spin briefly.
- 2. Add reaction mix to a TwistAmp® Basic reaction. Pipette to mix.
- 3. Add 2.5 µl of 280mM Magnesium Acetate (MgOAc) (supplied) and mix well to start reaction.

Note: RPA reactions start as soon as MgOAc is added.

- 4. Incubate at 39°C for 20 minutes. Remove strip after 4 minutes, vortex and spin briefly, replace in heating device.
- 5. After 20 minutes, clean amplicons before running on agarose gels.

Warning: If tubes are opened after amplification there is a great risk of contamination of work surfaces with amplicon. Ensure that appropriate avoidance measures are taken.

