Notice to purchaser

Absent a written agreement between you and TwistDx Limited to the contrary, the supply of this manual ("Manual") and the product described herein ("Product") is subject to the TwistDx Limited Terms and Conditions of Supply available at https://www.twistdx.co.uk/terms-of-supply, including, without limitation, the use restrictions, warranty disclaimer and limitation of liability set forth therein.

Use Restrictions

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

The Products are for non-commercial internal research use only. The Products must not be used for any *in vivo* testing, nor with humans or animals for any clinical diagnostic purposes. See the TwistDx Limited Terms and Conditions of Supply available at https://www.twistdx.co.uk/terms-of-supply for additional use restrictions.

No Warranty

TwistDx Limited makes no warranty with respect to the Product and excludes any warranties implied by law to the fullest extent permitted by law. THE PRODUCT IS PROVIDED WITHOUT ANY WARRANTY AS TO RESULTS, MERCHANTABILITY, FITNESS FOR A PARTICULAR PURPOSE OR NON-INFRINGEMENT OF ANY PATENT OR OTHER INTELLECTUAL PROPERTY RIGHT, AND WITHOUT ANY OTHER REPRESENTATION, WARRANTY OR CONDITION, EXPRESS OR IMPLIED, EXCEPT AS STATED IN THE PRECEDING SENTENCE. See the TwistDx Limited Terms and Conditions of Supply available at https://www.twistdx.co.uk/terms-of-supply for additional terms and conditions.

TwistDx™
Abbott House, Vanwall Business Park, Vanwall Road, Maidenhead, SL6 4XE, UK
E info@twistdx.co.uk | twistdx.co.uk

TwistDx™
Unwind DNA's possibilities

TwistAmp™ Liquid Basic

Quick Guide

Part number: INLQBAS
Revision 4

TwistAmp™ Liquid Basic Quick Guide

Please see instruction and assay design manuals at twistdx.co.uk for information regarding components and storage, assay design, and detailed use.

Instructions are based on 50 µl reaction volumes; if using a different volume, quantities should be adjusted appropriately.

Primer screen set-up (single-plex)[1]

1. Add 2.4 µl of each primer at 10µM concentration to 0.2 ml PCR tubes.

2. Prepare a pre-master mix (per reaction) in the order below:

2x Reaction Buffer	25 µl
dNTPs[2] + water[3] to	9.2 µl
10x Basic E-mix	5 µl

Vortex and spin briefly.

3. To the pre-master mix, add 2.5 µl 20x Core Reaction Mix[4] (per reaction) to tube lid. Mix by 10x full inversions and spin briefly. Master mix is now complete[5]. Pipette mix before use.

4. Add 41.7 µl[3] of master mix to primers prepared in tubes (step 1) and pipette mix.

5. Add 2.5 µl of 280mM MgOAc (supplied) and 1 µl template to tube lids[3]. DNA and MgOAc should be kept separate in the tube lid prior to spindown. Spin in MgOAc/template and mix well (6x inversions) to start reaction. Spin briefly.

Warning: RPA reactions start as soon as MgOAc is added.

6. Incubate at 37-42°C for 20-40 minutes. For low template copies, remove strip after 4 mins, mix by 6x full inversions and spin briefly, replace in heating device.

7. After step 6, clean amplicons before running on an agarose gel.

Warning: Opening tubes post amplification will risk contamination of work surfaces with amplicon. Ensure appropriate control measures are taken.

TwistAmp™ Liquid Basic Quick Guide

Template screen set-up (single-plex)[1]

1. Prepare a primer pre-master mix (per reaction) in the following order:

2x Reaction Buffer	25 µl
dNTPs[2] + water[3] to	9.2 µl
10x Basic E-mix	5 µl
Primer A (10µM)	2.4 µl
Primer B (10µM)	2.4 µl

Vortex and spin briefly.

2. Add 2.5 µl 20x Core Reaction Mix[4] (per reaction) to tube lid. Mix by 10x full inversions and spin briefly. Master mix is now complete[5]. Pipette mix before use.

3. Add 46.5 µl[3] master mix to 0.2 ml PCR tubes.

4. Add 2.5 µl of 280mM MgOAc and 1 µl template to tube lid[3]. DNA and MgOAc should be kept separate in the tube lid prior to spin-down. Spin in MgOAc/template, mix well (6x inversions) to start reaction. Spin briefly.

Warning: RPA reactions start as soon as MgOAc is added.

5. Incubate at 37-42°C for 20-40 minutes. For low template copies, remove strip after 4 mins, mix by 6x full inversions and spin briefly, replace in heating device.

6. After step 5, clean amplicons before running on an agarose gel.

Warning: Opening tubes post amplification will risk contamination of work surfaces with amplicon. Ensure appropriate control measures are taken.

1 See manual for multiplexing.
2 Suggested final concentration of 1.8mM (total) dNTPs. Optimisation is recommended.
3 Volumes should be adjusted if adding more/less template and/or MgOAc.
4 Warm to room temperature and pipette mix slowly to ensure homogeneity.
5 Master mix may appear cloudy, this is normal.

TwistDX
Liquid Basic QG

Size:
8.268 in x 5.827 in

PMS 185 C
Red

Black

PMS 7541 C
Gray 1 - 10%

Part Number: INLQBAS
Revision 4

Date of Revision:
4.2 2020/06/26